

RESEARCH

Open Access



Characterization of a novel *N*-acylhomoserine lactonase, AidP, from Antarctic *Planococcus* sp.

Wah Seng See-Too^{1,2}, Peter Convey^{2,3}, David A. Pearce^{2,3,4} and Kok-Gan Chan^{1,5*} 

Abstract

Background: *N*-acylhomoserine lactones (AHLs) are well-studied signalling molecules produced by some Gram-negative Proteobacteria for bacterial cell-to-cell communication or quorum sensing. We have previously demonstrated the degradation of AHLs by an Antarctic bacterium, *Planococcus versutus* L10.15¹, at low temperature through the production of an AHL lactonase. In this study, we cloned the AHL lactonase gene and characterized the purified novel enzyme.

Results: Rapid resolution liquid chromatography analysis indicated that purified AidP possesses high AHL-degrading activity on unsubstituted, and 3-oxo substituted homoserine lactones. Liquid chromatography–mass spectrometry analysis confirmed that AidP functions as an AHL lactonase that hydrolyzes the ester bond of the homoserine lactone ring of AHLs. Multiple sequence alignment analysis and phylogenetic analysis suggested that the *aidP* gene encodes a novel AHL lactonase enzyme. The amino acid composition analysis of *aidP* and the homologous genes suggested that it might be a cold-adapted enzyme, however, the optimum temperature is 28 °C, even though the thermal stability is low (reduced drastically above 32 °C). Branch-site analysis of several *aidP* genes of *Planococcus* sp. branch on the phylogenetic trees also showed evidence of episodic positive selection of the gene in cold environments. Furthermore, we demonstrated the effects of covalent and ionic bonding, showing that Zn²⁺ is important for activity of AidP in vivo. The pectinolytic inhibition assay confirmed that this enzyme attenuated the pathogenicity of the plant pathogen *Pectobacterium carotovorum* in Chinese cabbage.

Conclusion: We demonstrated that AidP is effective in attenuating the pathogenicity of *P. carotovorum*, a plant pathogen that causes soft-rot disease. This anti-quorum sensing agent is an enzyme with low thermal stability that degrades the bacterial signalling molecules (AHLs) that are produced by many pathogens. Since the enzyme is most active below human body temperature (below 28 °C), and lose its activity drastically above 32 °C, the results of a pectinolytic inhibition assay using Chinese cabbage indicated the potential of this anti-quorum sensing agent to be safely applied in the field trials.

Keywords: Quorum quenching, Episodic positive selection, *Pectobacterium*, Soft-rot disease

*Correspondence: kokgan@um.edu.my

¹ Division of Genetics and Molecular Biology, Institute of Biological Sciences, Faculty of Science, University of Malaya, 50603 Kuala Lumpur, Malaysia

Full list of author information is available at the end of the article



Background

Bacteria rely on quorum sensing (QS), or bacterial cell-to-cell communication, to coordinate gene expression at high cell densities [1]. *N*-acylhomoserine lactones (AHLs) are among the best-characterized chemical signalling molecules produced by a large number of Gram-negative Proteobacteria [2, 3]. AHL-regulated QS traits are initiated when the accumulation of AHLs achieves a critical threshold concentration with sufficient molecules present to bind on the cognate transcriptional regulators, to subsequently trigger the expression of the targeted gene [4]. QS regulates the expression of up to 10% of total Open Reading Frames (ORFs) in bacterial genomes [5] and more than 20% of the proteomes [6]. Thus, QS confers a significant advantage to bacterial populations, by allowing them to behave like an organized multicellular organism [7, 8].

AHL-mediated QS has been shown to be involved in the infection phenotype of *Pseudomonas aeruginosa* [9], *Aeromonas hydrophila* [10], *Pectobacterium carotovorum* subsp. *carotovorum* [11] and a wide range of other pathogens. Some bacteria produce AHLs to regulate phenotypes that cause detrimental effects within the environment. For example, the structure of the microbial community in corals infected with Black Band disease is associated with bacteria that possess AHL-mediated QS [12, 13]. Therefore, the disruption of QS, known as quorum quenching (QQ), is of interest for its potential application in mitigating the detrimental effects caused by AHL-producing pathogenic bacteria.

There are two families of AHL-degrading QQ enzymes, AHL-lactonases and AHL-acylases. AHL-lactonases catalyse AHL ring opening by hydrolysing lactones, and examples include AiiA from *Bacillus* sp. [14], AiiB and AttM from *Agrobacterium tumefaciens* [15, 16], and QsdA from *Rhodococcus erythropolis* [17]. AHL-acylases hydrolyse the amide bond of AHLs, and reported AHL-acylases included AiiD from *Ralstonia* sp. [18], PvdQ from *P. aeruginosa* [19] and AhlM from *Streptomyces* sp. [20]. These AHL-degrading QQ enzymes have been reported in bacteria, yeast [21], and mammalian sera [22], and the genes encoding these enzymes have also been found in a metagenomic library constructed from a soil sample [23]. AHL-oxidoreductases are another family of QQ enzymes produced by *R. erythropolis* that only modify the functional group of AHLs to affect the QS activity, unlike the other AHL-degrading enzymes [17]. Various plants have also been reported to produce QS inhibitors. For instance, halogenated furanones from *Delisea pulchra* inhibit QS-mediated gene expression by promoting the degradation of the bacterial transcriptional activator [24, 25].

Unlike other antibacterial strategies that employ antibiotics and which have led to the emergence of multidrug-resistant pathogens [26], QQ does not affect bacterial viability and therefore, imposes a lower selective pressure [27]. QQ has been employed in prevention of biofilm formation on membrane bioreactors used in wastewater treatment [28], biofouling in reverse osmosis (RO) systems [29] and for the degradation of aromatic compounds [30, 31]. This strategy has also been exploited by co-culturing the heterologous system of the QQ gene or purified QQ enzyme with AHL-producing pathogens, such as *P. carotovorum*, for plant disease control [14]. Dong et al. [14] also demonstrated that heterologous expression of *aiiA* in *P. carotovorum* significantly reduced the release of AHLs and attenuated the pathogenicity of the plant pathogen.

We have previously demonstrated that the bacterial strain *Planococcus versutus* L10.15^T, isolated from Antarctica, produces a novel class of AHL-lactonase. Homologous genes of *aidP* were identified in all type strains of *Planococcus* isolated from Antarctica, and we thus termed the gene encoding the AHL-lactonase as 'autoinducer degrading gene from *Planococcus* sp.' (*aidP*) [32]. In this study, we report the cloning of *aidP* from *P. versutus* L10.15^T and characterization of this novel AHL lactonase. We also further demonstrate that AidP is able to attenuate the virulence of *P. carotovorum* subsp. *carotovorum*, a plant pathogen that causes soft-rot disease in Chinese cabbage.

Materials and methods

Bacterial strains, plasmids, chemicals, and growth conditions

Bacterial strains and plasmids used in this study are listed in Additional file 1: Table S1. *Planococcus versutus* strain L10.15^T was isolated from a soil sample collected on Lagoon Island (Ryder Bay, Adelaide Island, maritime Antarctic), near to an elephant seal haul out and wallow area [32, 33]. *Escherichia coli* was grown at 37 °C in Luria–Bertani (LB) medium or FOC medium following the manufacturer's instructions (ChampionTM pET100 Directional TOPO[®] Expression Kit, Invitrogen). The AHL biosensor *Chromobacterium violaceum* CV026 [34] was maintained on LB agar at 28 °C, and was used to detect short-chain (C₄ to C₈) HSL. Antibiotic was added at a final concentration of 100 µg/ml for ampicillin or 50 µg/ml for kanamycin, as required. *P. versutus* strain L10.15^T was grown in LB medium at 16 °C. The AHLs used in this study, *N*-hexanoyl-L-homoserine lactone (C₆-HSL), *N*-heptanoyl-L-homoserine lactone (C₇-HSL), *N*-octanoyl-L-homoserine lactone (C₈-HSL), *N*-decanoyl-L-homoserine lactone (C₁₀-HSL), *N*-docecanoyl-L-homoserine lactone (C₁₂-HSL),

N-(3-oxohexanoyl)-*L*-homoserine lactone (3-oxo- C_6 -HSL), *N*-(3-oxooctanoyl)-*L*-homoserine lactone (3-oxo- C_8 -HSL), *N*-(3-oxodecanoyl)-*L*-homoserine lactone (3-oxo- C_{10} -HSL), and *N*-(3-oxododecanoyl)-*L*-homoserine lactone (3-oxo- C_{12} -HSL), were obtained from Sigma Aldrich (USA) or from Prof. Paul Williams (University of Nottingham). Isopropyl- β -D-1-thiogalactopyranoside (IPTG) was obtained from Promega (Madison, WI).

Molecular evolution analyses, amino acid profile analysis

All gene sequences were aligned in MEGA 7 [35] using MUSCLE [36]. The multiple sequence alignment analysis was conducted using Jalview version 2 [37] and highlighted according to Clustal X (refer to Additional file 1: Table S2 for colour code interpretation). In order to investigate the presence of selective forces acting on *aidP* genes, branch-site analysis for detection of positive selection using the CodeML program within PAML 4.7 [38] was performed. The ML phylogenetic tree inferred in the analysis was constructed using sequences obtained from a BLASTn search using the *aidP* gene sequence from *P. versutus* L10.15^T (WP_049694637.1) (Additional file 1: Figure S1). Likelihood ratio tests (LRT) were used to test for significance between the null and alternative models (χ^2 distribution, p value < 0.05). The Bayes empirical Bayes method (BEB) was used to identify positively selected sites when significant results were found [39]. The amino acid composition was calculated from the aligned region of gene sequences using CodeML analysis, and the heatmap was created using MORPHEUS (<https://software.broadinstitute.org/morpheus/>). The 3D structure of AidP was modelled using the automated SWISS-MODEL server [40]. Positively selected sites were highlighted using PyMOL.

Cloning of *aidP* gene

The full length *aidP* gene from *P. versutus* L10.15^T was amplified by PCR using primer pair PAL200Fa (5'-CAC CATGACTGGTATTATCAAGCC) and PAL200R (5'-TTATTTCGTAGTATCCTTCAGTCGACT). CACC, which pairs with the overhang sequence of the pET200 vector, was added at the 5' end of the forward PCR primer, to enable directional cloning. PCR was performed using KAPA HiFi polymerase (Kapa Biosystem, USA), with the following cycling parameters: 95 °C for 30 s, 62 °C for 30 s, and 72 °C for 1 min, for 35 cycles. The amplicons were purified using the AMPure XP-PCR purification system (Beckman Coulter, USA). The purified PCR product was cloned into the expression vector (pET-200) and transformed into *E. coli* TOP-10TM. Next, the pET200-*aidP* was amplified in the TOP-10TM host, and extracted using the Qiagen Plasmid Mini kit (Qiagen,

Germany). The pET200-*aidP* was then transformed into expression host *E. coli* BL21 StarTM.

Expression and purification of AidP protein

The gene was expressed in BL21 StarTM following the manufacturer's instructions (Invitrogen, USA). Briefly, BL21 StarTM harbouring pET200-*aidP* were grown in LB broth at 37 °C following the manufacturer's instructions (Invitrogen, USA). Protein expression was induced by the addition of 0.5 mM IPTG (isopropyl- β -D-thiogalactopyranoside). The cultures were further grown in 0.5 mM IPTG (150 rpm) for 16 h. Cells were harvested by centrifugation at 5000g for 10 min at 4 °C. The His₆-AidP protein was purified from the harvested cells using the Ni-NTA Fast Start Kit (Qiagen, Germany) under native conditions following the manufacturer's instructions. Briefly, the cells were lysed using the native lysis buffer provided by the kit, filtered using Fast Start Column, and finally eluted in a final buffer of 50 mM Na phosphate, 300 mM NaCl, and 250 mM imidazole, pH 8.0. The solubility of the protein was optimized using OptiSolTM Protein Solubility Screening kit (DiLYX, USA). Next, protein in buffer was dialyzed into 1 ml of reaction buffer [500 mM Na₂SO₄, and 1% Tween 20 (w/v), pH6]. The protein was normalized to 1 μ M and then purified by immunoprecipitation using Protein G Magnetic Beads (New England Biolabs, UK). The apparent molecular mass of the purified protein was determined by SDS-PAGE (12% [w/v]) after staining it and molecular mass standards with Coomassie brilliant blue R-250. Concentrations of the purified protein were routinely determined by the Bradford method [41], Qubit[®] 2.0 Fluorometer (Life Technologies, USA) and Bioanalyzer 2100 (Agilent Technologies, USA) with High Sensitivity Protein Screening Chip.

Detection of AHL-degrading activity of AidP

The mechanism of AidP AHL-degrading activity and chemical structure of products from the reaction between AidP and AHLs were determined. C_6 -HSL was first subjected to digestion by AidP, and the remaining C_6 -HSL was detected using AHL biosensor CV026, which produces purple pigment violacein in response to exogenous short chain AHLs [34]. The degradation of AHLs was further confirmed using rapid resolution liquid chromatography (RRLC, Agilent Technologies, USA). The mechanism of AidP catalysis was analysed using ultra performance liquid chromatography (UPLC) and electrospray ionization-mass spectrometry (ESI-MS). ESI-MS was performed with a 6460 triple-quadrupole instrument (Agilent Technologies, USA). Total of 1 μ M of AidP was mixed with 100 μ M of AHLs in 0.1 ml of reaction buffer. After 3 h incubation at 26 °C, the mixture was extracted three times with ethyl acetate, and the combined organic

phase was evaporated to dryness. The RRLC analysis was performed as previously described [42] with the samples being reconstituted in 0.1 ml ACN and separated using a symmetry EC-C₁₈ column (2.7 by 100 mm) (Agilent Poroshell 120 EC-C18). All experiments were conducted in triplicate, and all velocities were determined at time points by which no more than 20% of the substrates were consumed.

Characterization of enzymatic activity of AidP

The effects of various metal ions and the metal chelating reagent EDTA on AidP activity were examined both in vitro and in vivo. For in vitro assay, 1 mM EDTA or 1 mM concentration of each metal ion (Cu²⁺, Ca²⁺, Fe²⁺, Mn²⁺, Mg²⁺, Zn²⁺, or Co²⁺) were mixed individually with 1 μM of purified AidP protein in a reaction buffer containing 3-oxo-C₆-HSL. After incubation at 26 °C for 3 h, the remaining 3-oxo-C₆-HSL was measured using RRLC as described above. For in vivo assay, an overnight culture of *E. coli* harbouring the plasmid pET200-aidP was diluted into fresh LB medium with 1 mM EDTA or 1 mM concentration of each selected metal ion. 3-oxo-C₆-HSL and IPTG were added to induce AidP over-expression after 6 h cultivation. After an additional 6 h cultivation, crude cell extracts were prepared and adjusted to the same concentration before being subjected to RRLC analysis.

Inhibition of pectinolytic activity of *Pectobacterium*

carotovorum subsp. *carotovorum* strain GS101 and PNP22

The assay was performed Chinese cabbage as described by Lojkowska et al. [43] and Uroz et al. [44]. Plant material was gently washed with sterile H₂O, wiped with sodium hypochlorite, and then dried under sterile conditions. *P. carotovorum* wild type strain GS101 was used in this assay, with *luxI* mutant *P. carotovorum* GS101, strain PNP22 grown without 3-oxo-C₆-HSL as a negative control, or with 3-oxo-C₆-HSL at a final concentration of 1 μM as a positive control. Briefly, strains were grown overnight at 25 °C in LB medium, suspended and diluted in PBS. The bacterial suspensions were introduced into the plant material using sterile pipette tips as (1) GS101 with or without AidP enzyme to determine the effect of AidP to attenuate the pathogenicity of the wild type *P. carotovorum*, (2) PNP22 grown with or without 3-oxo-C₆-HSL to determine the roles of *luxI* gene in pectinolytic activity, and (3) PNP22 grown with 3-oxo-C₆-HSL, added with AidP enzyme, to further confirms that AidP attenuate the pathogenicity via degradation of the autoinducer. The plant material was incubated at 25 °C and 90% relative humidity for 5 days. The results were assessed by direct visual inspection.

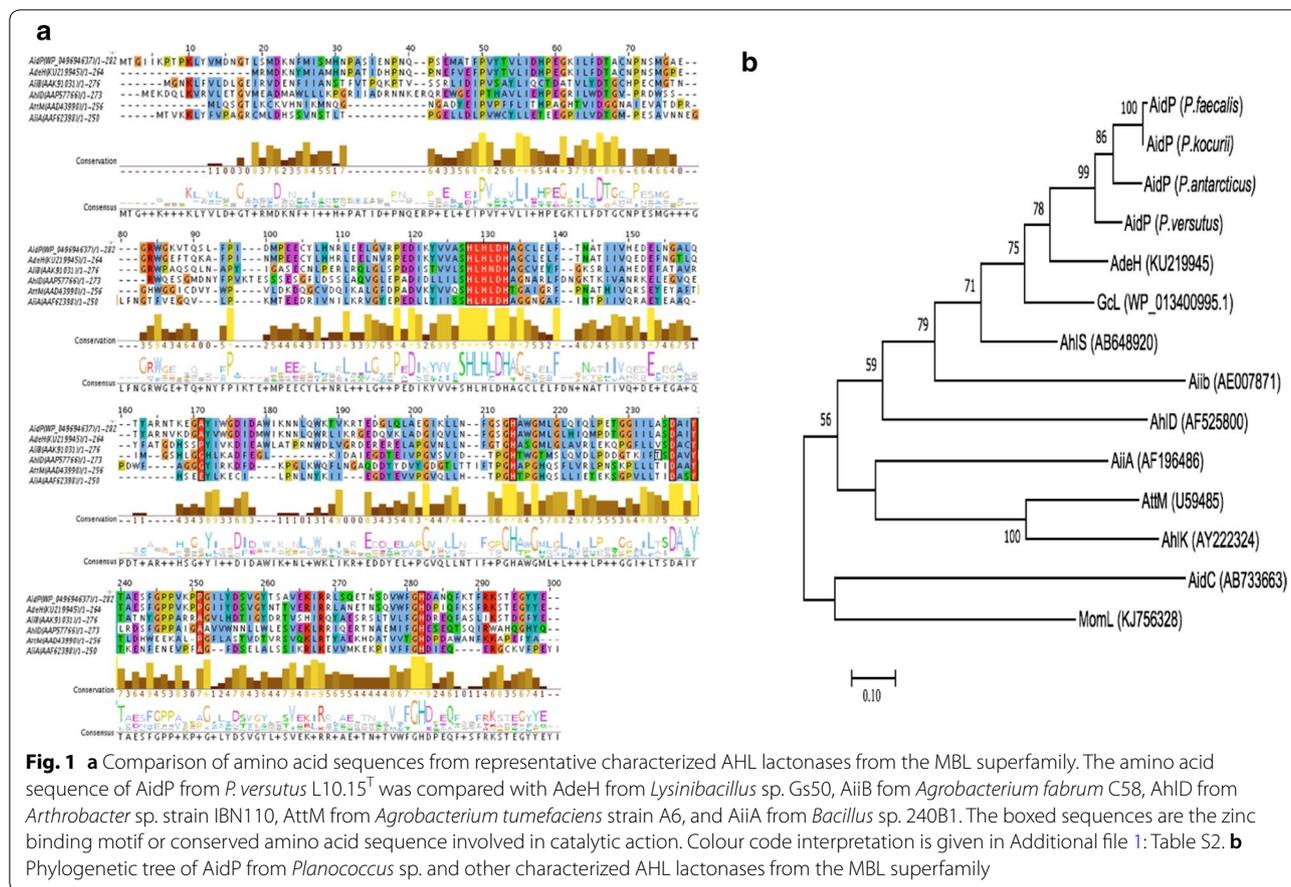
Results

Sequence analysis of AidP

From the multiple sequence alignment analysis, we have identified the HXHXDH~H zinc-binding motifs, which correspond to ¹¹⁷HLHLDH¹²²~H¹⁹⁷ in AidP, and also several functionally important amino acids based on a previous crystallographic study of AiiA-type AHL lactonase [45, 46] (Fig. 1a). These amino acids include D¹²¹ and D²¹⁹, which are important in forming bridging with the ligands, H²⁶⁵ and H¹²² that interact with the zinc metal ion and the active-site residue, Y²²², as an H⁺ donor. However, some amino acid substitutions were observed in the AidP. L¹²⁰ and A¹⁵⁶ are two amino acids that are substituted for F¹⁰⁷ and E¹³⁶ in AiiA AHL-lactonase, which form hydrogen bonds with the substrate and facilitate the interaction of the substrate with water molecule. One amino acid that is involved in the catalytic action (P²⁰³), in AidC and is involved in forming the structure of the protein was substituted by E¹⁸⁶ in AidP (Fig. 1a).

The phylogenetic analysis with all the known AHL lactonases from the MBL superfamily indicated that AidP is closely related to AdeH from *Lysinibacillus* and GcL from *Geobacillus* (Fig. 1b). GcL is a thermostable AHL-lactonase that is active at high temperature. However, phylogenetic analysis conducted using the NCBI BLAST search result indicated that the *aidP* gene is closely related to other hypothesized AHL lactonase gene from *Staphylococcus* spp. and *Sporosarcina ureae* (Additional file 1: Figure S1). These genes have yet to have their function characterized (in coding for AHL lactonases). Even though phylogenetic analysis on known AHL lactonases indicates that AidP has a close phylogenetic relationship with a thermophilic enzyme, amino acid substitutions at crucial sites can fundamentally change the flexibility of proteins. This also leads to some common characteristics in the amino acid composition of cold-adapted enzymes. Therefore, we conducted amino acid usage analysis on AidP and the nearest homologous genes including the thermophilic counterpart of this enzyme. The amino acid usage analysis indicated that AidP has high frequencies of Gly (G), Ser (S), Thr (T), and Leu (L) and low frequencies of Arg (A), Phe (F), and Met (M) compared to the nearest homologous genes, including the thermally-stable AHL-lactonase obtained from thermophilic bacteria such as *Geobacillus*, *Parageobacillus* and *Thermaerobacter* (Fig. 2, Additional file 2: Table S3).

Since the results of phylogenetic analyses showed that AidP genes form a hitherto unknown subline in the phylogenetic trees, we hypothesized that there is selective pressure may have acted on AidP, leading to adaptation to cold environments. To find evidence of this selective pressure, we performed a branch-site analysis using CodeML, and inference was made from a phylogenetic



tree constructed from the nearest gene to AidP identified in an NCBI BLAST search (Additional file 1: Figure S1). Using a branch containing four *aidP* genes from different species of *Planococcus* as foreground branch, a significant result from the likelihood ratio test ($p < 0.05$; Table 1) was obtained. A total of 11 sites under positive selection pressure were identified in Bayes Empirical Bayes analysis. One of the sites had posterior probability > 0.95 , site 84S, and there were two glycine residues distant from the active site that had high posterior probability, sites 104G (0.885) and 183G (0.948) (Fig. 3). Our analysis suggested that AidP from the *Planococcus* sp. branches could be under episodic selective pressure potentially driving adaptation, since these *Planococcus* sp. strains were isolated from Antarctica [32].

Purification of AidP and determination of enzyme activity

The purified AidP was subjected to Bioanalyzer analysis using a high sensitivity protein assay, revealing that the protein was approximately 33 kDa in size (Fig. 4), consistent with the predicted molecular weight based on the amino acid sequence. When the AHL-degrading activity of the purified protein was examined, it completely

degraded 100 μ M 3OC₆-HSL within 24 h. Results from RRLC and MS analyses indicated that AidP is an AHL-lactonase that hydrolyses the lactone ring of AHLs to acylhomoserine lactone. The RRLC profile of C₆-HSL was characterized by a single peak with a retention time of 1.1 min. After being treated with AidP, a decrease in the milli-absorbance unit (mAU) indicated the AHL-degrading activity of AidP (Fig. 5a). ESI-MS analysis of this product revealed an (M+H) ion at an m/z of 218 (Fig. 5b), indicating the effect of AidP on C₆-HSL, causing a mass increase of 18, corresponding to the addition of a water molecule. These results suggested that the treatment of AHL with AidP led to the cleavage of the homoserine lactone ring on the substrate, thus producing an N-3-oxo-hexanoyl-L-homoserine molecule.

Substrate specificity and properties of AidP

To assess the AHL substrate range of AidP, the residual AHLs present after incubation with AidP were assessed by RRLC. AidP exhibited high relative activities toward 3-oxo substituted AHLs. Nonetheless, AidP exhibited wide activities toward most AHLs tested including those with 3-hydroxy substitution, and un-substituted

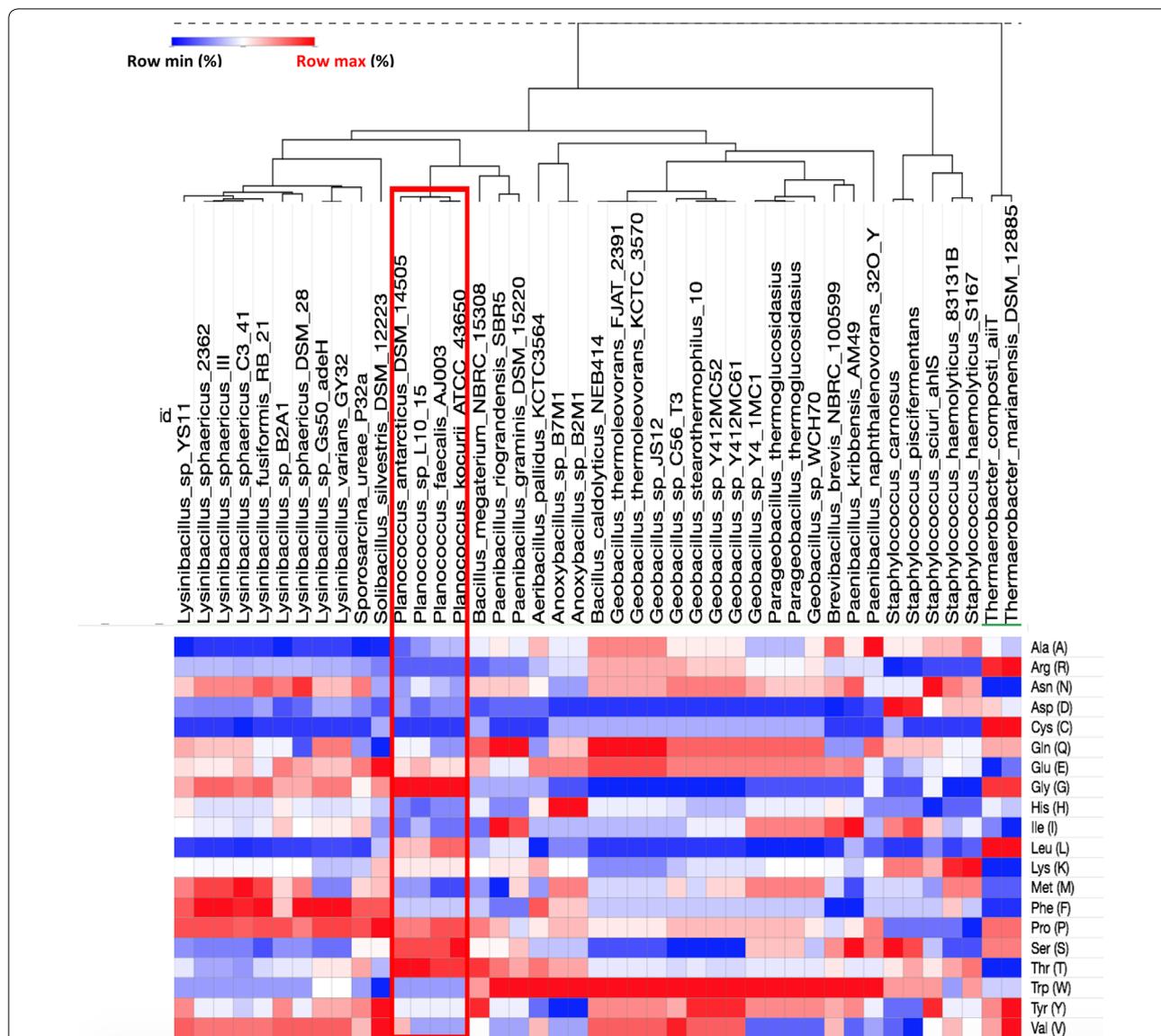


Fig. 2 Amino acid usage analysis based on aligned region of AidP gene (box) from *Planococcus* sp. and the nearest homologous gene based on NCBI BLAST result

homoserine lactone. AidP substrate specificity was significantly affected by the length of the acyl acid chain. AidP was comparatively effective in degrading C₆-C₁₀-HSL (Fig. 6a) but showed relatively low activity towards C₄-HSL and C₁₂-HSL.

The optimal pH for AHL-degrading activity of AidP was examined using 3OC₆-HSL as the substrate at 25 °C. AHL-degrading activity was enhanced as pH increased and reached a maximum at pH 8, while no or little activity was detected when pH was adjusted to pH 4 or lower (Fig. 6b). To determine the thermal stability of AidP, the enzyme was pre-incubated at various

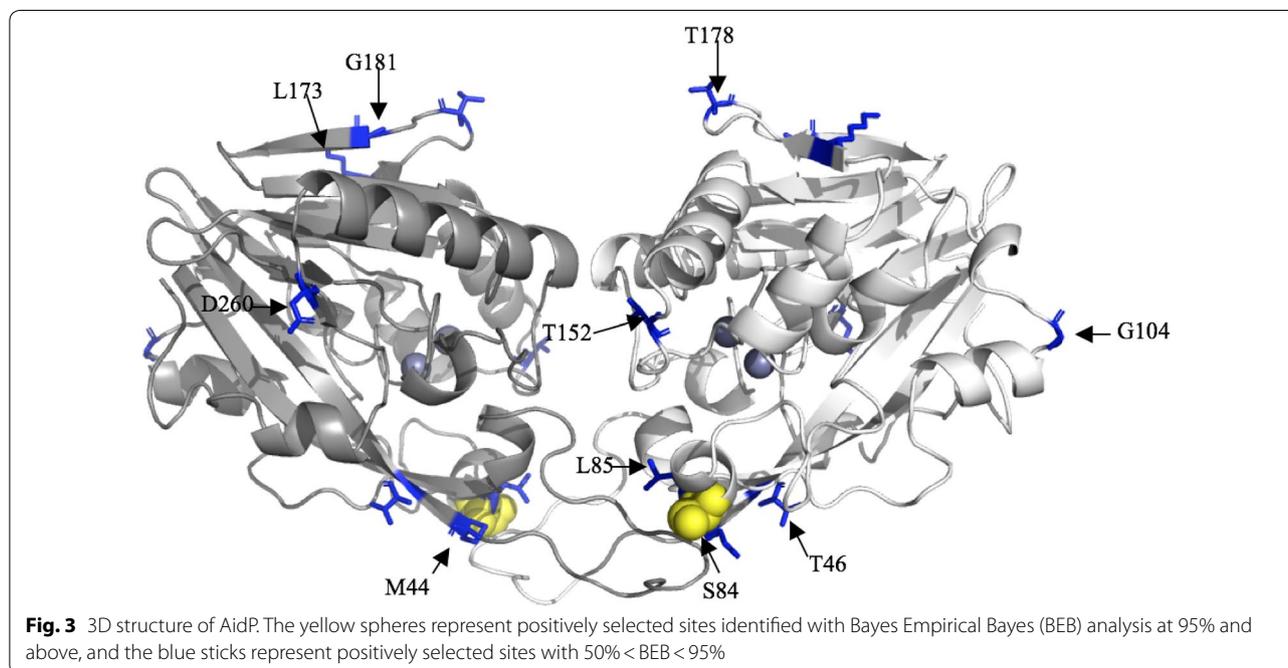
temperatures for 3 h and the residual enzymatic activity was then determined. The optimal temperature for the AHL-degrading activity of AidP was also examined using 3OC₆-HSL as the substrate, and was found to be approximately 27 °C, with activity decreasing at temperatures higher than 30 °C (Fig. 6c). The relative activity of AidP at 4 °C was less than 10% of the maximum value, but it increased to 60% at 16 °C. More than 90% of the activity remained after incubation at temperatures of 30 °C or less, although activity was markedly reduced after incubation at temperatures over 32 °C (Fig. 6d).

Table 1 Summary of CodeML results for branch-site analyses for *aidP* genes and 42 homologous gene obtained from NCBI BLAST result

Model	Site class	Proportion	Background ω	Foreground ω	<i>InL</i>	LRT Statistic	* <i>p</i> -value
Branch-site Null	0	0.89838	0.02528	0.02528	- 11,269.67116	4.42473	0.035422
	1	0.01171	1.000	1.000			
	2a	0.08875	0.02528	1.000			
	2b	0.00116	1.000	1.000			
	Branch-site Alternative Model	0	0.91669	0.02528			
	1	0.01197	1.000	1.000	- 11,267.458798		
	2a	0.07041	0.02528	38.61239			
	2b	0.00092	1.000	38.61239			

Foreground branch contains *aidP* genes for four *Planococcus* spp.

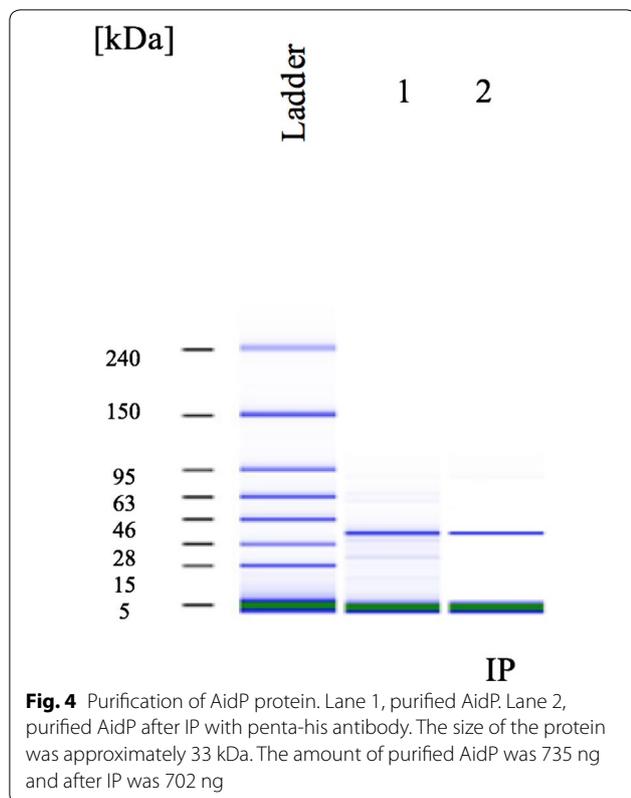
**p*-value was determined using χ^2 analysis with degree of freedom = 1



Zinc is essential for the AHL-degrading activity of AidP

Sequence alignment of AidP with the known AiiA-type lactonases revealed that they share the zinc-binding motifs HXHXDH~H (Fig. 1a). To verify whether AidP requires any cofactor for its enzymatic activity, the effects of several divalent metal ions on the AHL-degrading activity of AidP were evaluated. For in vitro assay, purified AidP was mixed with 3OC₆-HSL in reaction buffer containing either EDTA or metal ions. When 1 mM EDTA was added to this reaction mixture, the AHL-degrading activity of AidP decreased by approximately 80% (Fig. 6d). AidP activity was also markedly inhibited by the addition of Mg²⁺. The reported metal ion inhibitor Fe²⁺ [51] showed no significant inhibitory effect on AidP.

AidP activity was successfully restored to 85% of the original level by the subsequent addition of Zn²⁺, but not by any of the other metal ions tested (Fig. 6e). For in vivo assays, *E. coli* harbouring pET200-aidP was cultivated in LB medium containing EDTA or metal ions, and the AHL-degrading activity of AidP in the crude cell extracts was then determined. The AHL-degrading activity of AidP was extinguished through the addition of 1 mM EDTA but was again restored by the addition of Zn²⁺ (Fig. 6f). Furthermore, most other metal ions showed very low inhibitory effect on enzyme activity, with activity being reduced by at most 30%, with the exception of Cu²⁺, which reduced activity by more than 50%. No other metal ion stimulated AidP activity.



AidP attenuated QS-regulated functions of plant pathogenic bacteria

The potential of AidP as an anti-QS reagent was assessed in a pectinolytic inhibition assay with Chinese cabbage. After 3 days incubation at high humidity, maceration of tissue was observed in cabbages inoculated only with GS101 (Fig. 7a). When GS101-inoculated plant material was incubated together with AidP, the area of tissue maceration was significantly reduced, and maceration was totally prevented with 10 μ M AidP (Fig. 7a). The *luxI* mutant PNP22 had completely lost the pectinolytic activity (Fig. 7b), whereas the complementation of the mutant strain with exogenous AHL (3-oxo-C₆-HSL) successfully restored pectinolytic activity (Fig. 7b). The effect of AidP in the complementation assay, was further investigated, with the data indicating that AidP had significantly reduced the pectinolytic activity of PNP22 complemented with exogenous AHL (Fig. 7b). These observations suggest that AidP is able to disrupt the QS signal from the pathogenic bacteria and hence has the potential to act as a biocontrol agent.

Discussion

In this study, a novel AHL lactonase, AidP, was identified and characterized from the type strain of *P. versutus*. AidP is also the first reported AHL-lactonase

enzyme from a bacterium isolated from Antarctica. We classified AidP as a novel AHL lactonase because of its low sequence identity compared with other AHL lactonases. Previously, we have shown that AHL-degrading *P. versutus* L10.15^T exhibits AHL-degrading activity at temperatures as low as 4 °C [32]. The data obtained in the current study further suggested that AidP may have structural modifications that enable its maintenance of high activity at low temperatures, thereby extending the known diversity of AiiA-type lactonases.

Amino acid composition and substitution analyses of cold-adapted enzymes have been conducted by numerous groups, to examine the structural flexibility of these enzymes at low temperature [47, 48]. A single substitution of an amino acid can stabilize/destabilize the protein structure due to the side-chain functional group of the amino acid, and drastically affect the thermal stability and thermodynamics of the protein [49, 50]. These studies have identified a number of prominent characteristics in cold-adapted enzymes, consistent with our observations on AidP, in particular having the (1) highest glycine and (2) lowest arginine composition amongst the closely related thermophilic and mesophilic homologous genes (Fig. 2). Glycine that is not in the active site (as the active site is always conserved), substantially contributes to the flexibility of the cold-adapted enzyme at low temperature [51]. Cold-adapted enzymes were observed to have low arginine composition, as arginine provides surface salt bridges or ion pairs bonding secondary structures in protein domains [52, 53]. Hawwa et al. [54] have proposed that high thermal stability in phosphotriesterase-like AHL lactonases is due to the reduced number of glycine residues in this family of AHL lactonases. *P. versutus* L10.15^T was originally isolated from Antarctic soils and we have previously demonstrated that it is capable of degrading AHLs at temperatures as low as 4 °C [32]. At low temperature, decrease in pH could affect the charge of amino acids, particularly histidine residues [55]. Thus, the apparent loss of efficiency in the isolated enzyme at 4 °C compared to the high degradation activity recorded from entire bacterial cells could indicate that the zinc-binding motifs with high histidine (HXHXDH~H), that are important for structural stabilization and catalysis, have been affected [56]. In the bacterial cells, we speculate there are discrete structural modifications which allow the protein to remain active at low temperatures in the living cell (Fig. 6e, f).

Phylogenetic analysis of AidP along with other known AHL-lactonases from the MBL superfamily indicated that AidP is closely related to AdeH from *Lysinibacillus* and GcL from *Geobacillus*. AdeH has a similar active temperature range as AidP, with enzyme activity significantly reduced above 30 °C. AdeH was identified

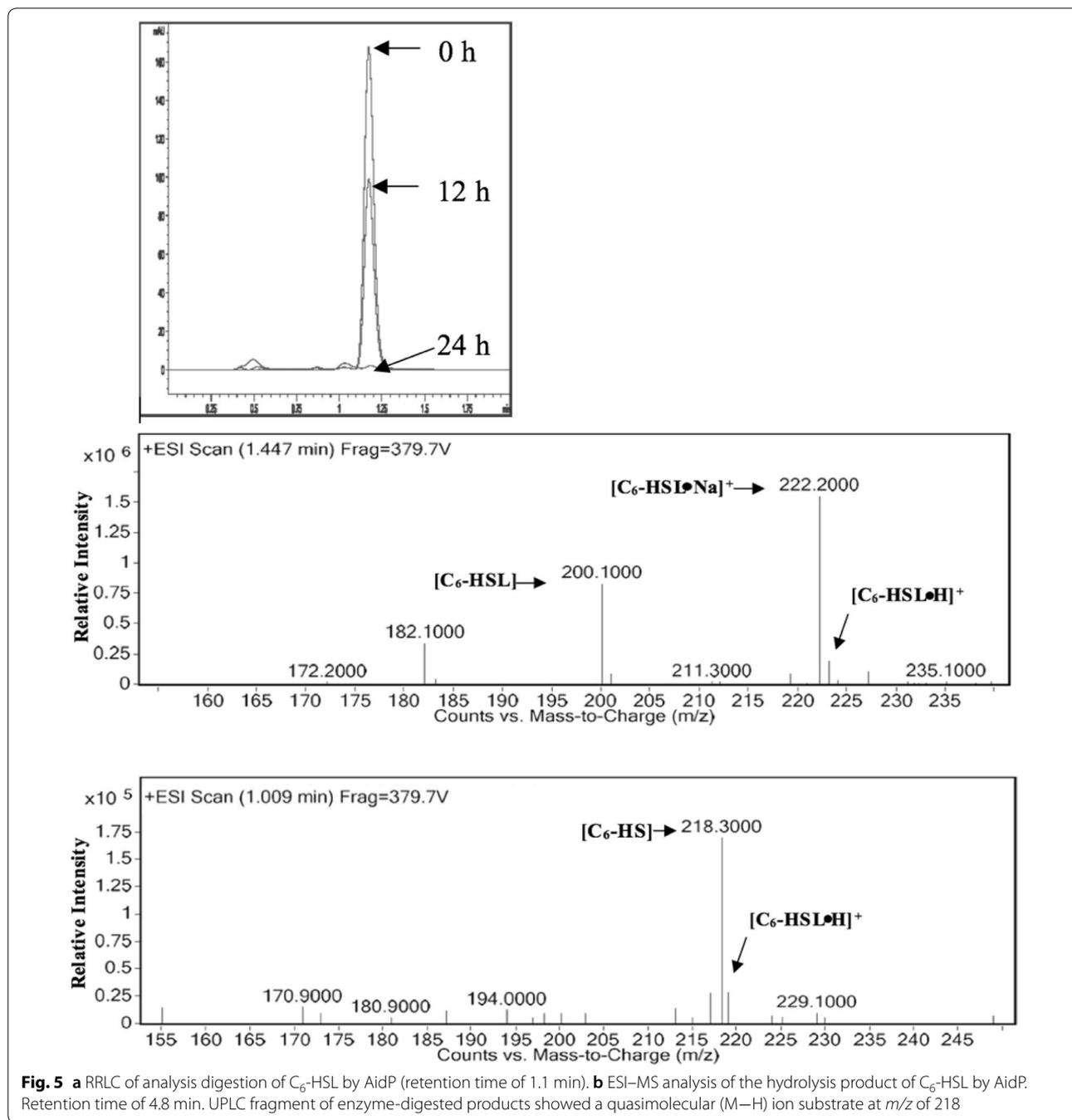
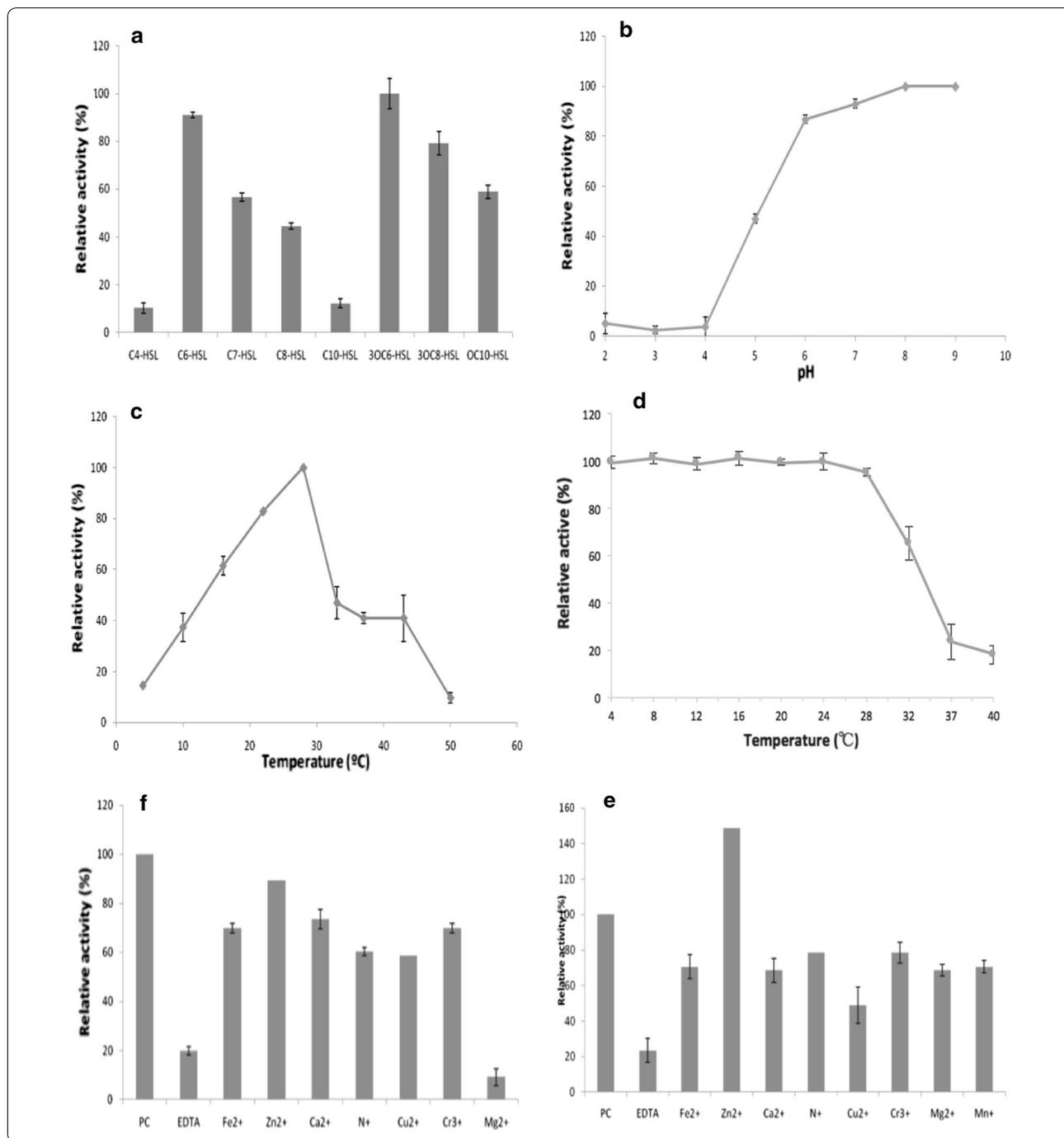


Fig. 5 a RRLC of analysis digestion of C₆-HSL by AidP (retention time of 1.1 min). b ESI-MS analysis of the hydrolysis product of C₆-HSL by AidP. Retention time of 4.8 min. UPLC fragment of enzyme-digested products showed a quasimolecular (M-H) ion substrate at m/z of 218

in *Lysinibacillus* isolated from soil, which is the most closely related genus to *Planococcus* that possesses AHL lactonase genes. Another closely related AHL lactonase, GcL isolated from the genus *Geobacillus*, is also closely related to *Planococcus*. However, it is a thermostable protein that is active at a high optimum temperature. These AHL lactonase genes appear to be closely related to each other with recent divergences. Variations

in amino acid composition among these enzymes may explain the difference in their thermal stability. The result also prompted us to carry out branch-site analysis to test for positive selection of *aidP* genes. As the closely related genes are so diverse in terms of active temperature range, we suggest that there is selection pressure on the gene, that tunes the enzyme entropy in order to function well in cold environments. Making



inference from the phylogenetic tree of these genes, we found several sites in AidP that could be under positive selection, indicating that the protein might had experienced episodic selection pressure in cold environments. Saavedra et al. [57] demonstrated how glycine substitution distant from the active-site affected the thermodynamics of a cold-adapted enzyme. This may also explains

why AidP, while being closely related to a thermophilic enzyme, has distinct thermodynamic and thermal stability characteristics.

AidP exhibited the lowest optimum temperature among all the closely related AHL lactonases. Most of the AHL-lactonase genes previously identified in different taxa have optimum temperature from 20 to 50 °C,

(See figure on previous page.)

Fig. 6 **a** Substrate specificity of purified AidP. Purified AidP was mixed with substrate solutions in buffer (pH 7). After incubation at 25 °C for 30 min, the residual AHL and its hydrolysis products were quantified by RRLC. We defined 100% relative activity as the activity toward 3OC₆-HSL. Each incubation was replicated at least three times, and error bars indicate standard deviation. **b** Optimal pH of the AHL-degrading activity of purified AidP. Purified AidP was mixed with 3-oxo-C₆-HSL in reaction buffers with pH ranging from pH 2 to pH 9 and incubated at 25 °C. Each incubation was replicated at least three times, and error bars indicate standard deviation. **c** Optimal temperature of AHL-degrading activity of purified AidP. Purified AidP was mixed with 3OC₆-HSL in reaction buffer (pH 7) and incubated at temperatures ranging from 4 to 50 °C. After incubation for 30 min, the residual substrate was quantified by RRLC. We defined 100% relative activity as the activity in the reaction buffer (pH 7) at 25 °C. Control samples, which had been prepared under the same conditions but without AidP, were used to exclude the influence of autodegradation at high pH or high temperature. Each incubation was replicated at least three times, and error bars indicate standard deviation. **e** Thermal stability of AidP. Purified AidP reaction buffer (pH 7) was incubated at temperatures ranging from 4 to 40 °C for 30 min before subjected to AHLs inactivation assay with 3-oxo-C₆-HSL. After incubation for 30 min, the residual substrate was quantified by RRLC. We defined 100% relative activity as the activity in the reaction buffer (pH 7) at 25 °C. Each incubation was replicated at least three times, and error bars indicate standard deviation. The effects of EDTA and various metal ions on the AHL-degrading activity of AidP in in vitro (**e**) and in vivo (**f**) assays. For the in vitro assay, purified AidP was incubated in reaction buffer containing substrate and a 1 mM concentration of the appropriate metal ion. After incubation at 25 °C for 30 min, the residual activity of AidP was analyzed by RRLC. For the in vivo assay, *E. coli* BL 21 Star™ harboring pET200-aidP was cultivated with 1 mM EDTA and a 1 mM concentration of the appropriate metal ion. Crude cell extracts were then prepared and used for the AHL-degradation assay. We defined 100% relative activity as the activity in the absence of EDTA or metal ions, and represented the PC (positive control). Each incubation was replicated at least three times, and error bars indicate ± 1 standard deviation

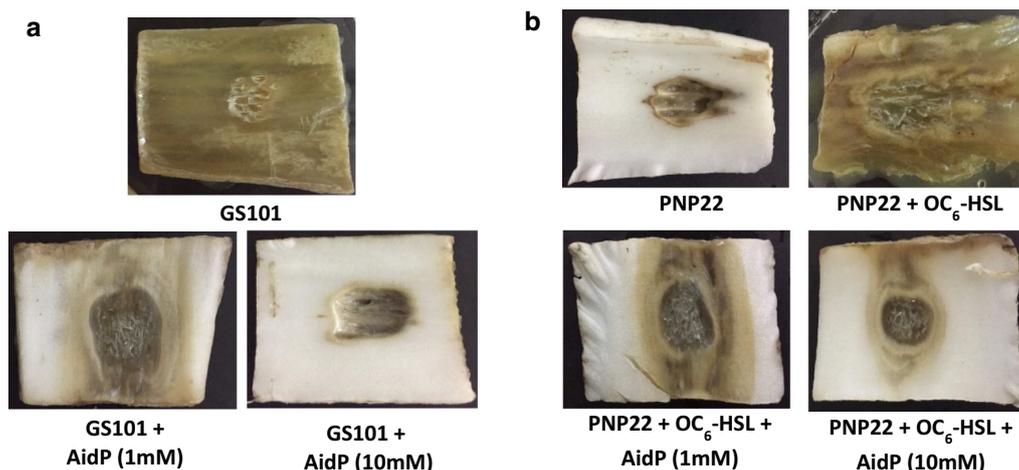


Fig. 7 Quenching of pectinolytic activity in *P. carotovorum* strain GS101 and PNP22 by AidP. The pectinolytic inhibition assay was observed by visual inspection of maceration zones by the pathogen *P. carotovorum* strain **a** GS101 and **b** *luxI* mutant PNP22 after 5 days

for example MomL [58], AidC [59], and AiiK [60]. AdeH, which is closely related to AidP, has a lower optimum temperature (37 °C) compared to other AHL lactonases. AidP is closely related to a number of thermally stable AHL lactonases, including GcL [61] and AiiT [62], which retains its activity at 70 °C and above.

Most known AHL lactonases, such as AiiA from *Bacillus* sp. 240B1 [63], show relatively high activity toward unsubstituted AHLs. Therefore, the high relative activities of AidP toward 3-oxo substituted AHLs compared to unsubstituted AHLs contrasts with characteristics previously reported for AHL lactonases, except AidC [59] (Fig. 6a). In contrast to MomL that has higher preference for long chain AHLs as substrate [58], AidP is similar in substrate specificity as AiiK [60], with higher activity

against mid-long chain AHLs (Fig. 6a). AidP exhibited high activity toward most AHLs tested, including those with 3-hydroxy substitutions. However, in common with AiiA, AidP did not show any degrading activity against L-homoserine lactone or γ -butyrolactone.

The increase in AHL-degrading activity of AidP achieved by the addition of Zn²⁺ suggests that this ion is essential for the AHL-degrading activity of AiiA-type lactonase. The inhibition of the AHL-degrading activity of AidP by the addition of Cu²⁺ indicated that this ion is an AHL lactonase inhibitor for AidP, as has previously been reported for other AHL lactonases [63]. Other known AHL lactonases from the MBL-superfamily, including AiiA [56], AiiB [64] and MomL [58], can bind to other metal ions including Mg²⁺, which in contrast had an inhibitory effect in the

in vitro assay for AidP. Nonetheless, AidP only binds to Zn^{2+} of all the metal ions tested. The effect of these metal ions on AidP is similar to those on its nearest known AHL lactonase neighbour, AdeH [65].

Most studies of AHL lactonases to date have focussed on the human pathogen, *Pseudomonas aeruginosa*. Migiyama et al. [66] has demonstrated that AHL lactonase AiiM is effective against *P. aeruginosa* in the mouse model of acute pneumonia. Tang et al. (2015) [58], on the other hand, demonstrated that MomL is effective against *P. aeruginosa* in *Caenorhabditis elegans*. AHL-lactonases such as AhLS [67, AidH 68] and AdeH [65] have also been proven effective in reducing pathogenicity of *P. carotovorum*, a species that causes soft-rot disease in various plants. When a gene encoding AHL-lactonase was transformed into *P. carotovorum*, AHL production was significantly reduced and the genes responsible for pectinolytic activity were not expressed [14, 69, 70]. A transgenic plant with the gene encoding AHL-lactonase also had substantially enhanced resistance against *P. carotovorum* [71]. These findings indicate the potential of AHL lactonases to be used as microbial antagonists.

The scientific community has however, raised concerns about the approaches used to create disease-resistant transgenic plants or crops. Firstly, resistance may be induced and developed in target bacteria even though AHL lactonases impose lower selection pressure compared to antibiotics [72]. Even though the effect of AHL lactonase is specific and potent [73], constant expression of the AHL lactonase in a plant or crop might also accelerate the emergence of resistance in the pathogen. Secondly, AHLs play crucial roles in plant defence responses, growth and development [74–76]. Therefore, expression of AHL lactonase in plant cells may also impact positive effects derived from bacteria-plant interactions. Thirdly, degradation of AHLs within plant cells itself may affect beneficial endophytes which possess AHL-mediated QS systems. Endophytes are known to improve plant rooting and nutrient uptake and enhance chlorophyll production [77]. Some endophytes produce AHLs to enhance their colonization on host plants and to counter phytopathogens [78]. Therefore, a strategy using direct application of AHL lactonases to target specific plant pathogens that produce AHLs could be recommended as treatment of infected plants, mitigating the dependency on antibiotics that could accelerate the development of antibiotic-resistant pathogens.

Conclusions

In this study, a novel AHL lactonase from *P. versutus* was successfully cloned and expressed. It showed considerable hydrolytic activity, and we demonstrated the ability to

attenuate pathogenicity of the plant pathogen *P. carotovorum* in vivo. We conclude that this anti-quorum sensing agent can be safely applied in field tests with the low thermal stability.

Additional files

Additional file 1: Figure S1. Phylogenetic tree constructed using NCBI BLAST result of *aidP* gene from *P. versutus* L10.15^T. The foreground branch is highlighted in red colour. **Table S1.** Bacterial strains and plasmids.

Table S2. Colour coding for multiple sequences alignment analysis.

Additional file 2: Table S3. Amino acid usage analysis based on aligned region *AidP* gene and the nearest homologous gene based on NCBI BLAST result.

Authors' contributions

WSST conceived the study and conducted the experiments. WSST, PC, DAP and KGC analysed the data and prepared the manuscript. All authors read and approved the final manuscript.

Author details

¹ Division of Genetics and Molecular Biology, Institute of Biological Sciences, Faculty of Science, University of Malaya, 50603 Kuala Lumpur, Malaysia.

² National Antarctic Research Centre, IPS Building, University Malaya, 50603 Kuala Lumpur, Malaysia. ³ British Antarctic Survey, NERC, High Cross, Madingley Road, Cambridge CB3 0ET, UK. ⁴ Applied Sciences, University of Northumbria at Newcastle, Newcastle-upon-Tyne NE1 8ST, UK. ⁵ International Genome Centre, Jiangsu University, Zhenjiang, China.

Acknowledgements

We thank Dr. Romain Studer for his assistance in branch-site analyses. Wah-Seng See-Too thanks the Bright Sparks Program of the University of Malaya for scholarship awarded.

Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

All data generated or analysed during this study are included in this published article.

Consent for publication

Not applicable.

Ethics approval and consent to participate

Not applicable.

Funding

This work was supported by University of Malaya Research Grants (PG203-2016A, GA001-2016), University of Malaya High Impact Research Grants (UM-MOHE HIR Grant UM.C/625/1/HIR/MOHE/CHAN/14/1, Grant No. H-50001-A000027; UM-MOHE HIR Grant UM.C/625/1/HIR/MOHE/CHAN/01, Grant No. A-000001-50001) awarded to Kok-Gan Chan. Peter Convey is supported by NERC core funding to the British Antarctic Survey's 'Biodiversity, Evolution and Adaptation' Team, and by a University of Malaya Visiting Icon Professorship.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 3 May 2018 Accepted: 7 November 2018

Published online: 17 November 2018

References

- Miller MB, Bassler BL. Quorum sensing in bacteria. *Annu Rev Microbiol*. 2001;55:165–99.
- Deep A, Chaudhary U, Gupta V. Quorum sensing and bacterial pathogenicity: from molecules to disease. *J Lab Physicians*. 2011;3:4–11.
- Galloway WR, Hodgkinson JT, Bowden SD, Welch M, Spring DR. Quorum sensing in Gram-negative bacteria: small-molecule modulation of AHL and AI-2 quorum sensing pathways. *Chem Rev*. 2010;111:28–67.
- Taga ME, Bassler BL. Chemical communication among bacteria. *Proc Natl Acad Sci USA*. 2003;100:14549–54.
- Wagner VE, Bushnell D, Passador L, Brooks AI, Iglewski BH. Microarray analysis of *Pseudomonas aeruginosa* quorum sensing regulons: effects of growth phase and environment. *J Bacteriol*. 2003;185:2080–95.
- Arevalo-Ferro C, Hentzer M, Reil G, Görg A, Kjelleberg S, Givskov M, Riedel K, Eberl L. Identification of quorum sensing regulated proteins in the opportunistic pathogen *Pseudomonas aeruginosa* by proteomics. *Environ Microbiol*. 2003;5:1350–69.
- Chhabra SR, Hart C, Hooi DSW, Daykin M, Williams P, Telford G, Pritchard DI, Bycroft BW. Synthetic analogues of the bacterial signal (quorum sensing) molecule *N*-(3-oxododecanoyl)-L-homoserine lactone as immune modulators. *J Med Chem*. 2003;46:97–104.
- Atkinson S, Williams P. Quorum sensing and social networking in the microbial world. *J R Soc Interface*. 2009;6:959–78.
- Lee J, Zhang LH. The hierarchy quorum sensing network in *Pseudomonas aeruginosa*. *Protein Cell*. 2015;6:26–41.
- Bi ZX, Liu YJ, Lu CP. Contribution of *AhyR* to virulence of *Aeromonas hydrophila* J-1. *Res Vet Sci*. 2007;83:150–6.
- Lee DH, Lim J, Lee J, Roh E, Jung K, Choi M, Oh C, Ryu S, Yun J, Heu S. Characterization of genes required for the pathogenicity of *Pectobacterium carotovorum* subsp. *carotovorum* Pcc21 in Chinese cabbage. *Microbiology*. 2013;159:1487–96.
- Zimmer BL, May AL, Bhedi CD, Dearth SP, Prevatte CW, Pratte Z, Campagna SR, Richardson LL. Quorum sensing signal production and microbial interactions in a polymicrobial disease of corals and the coral surface mucopolysaccharide layer. *PLoS ONE*. 2014;9:e108541.
- Meyer JL, Gunasekera SP, Scott RM, Paul VJ, Teplitski M. Microbiome shifts and the inhibition of quorum sensing by Black Band Disease cyanobacteria. *ISME J*. 2015;10:1–13.
- Dong YH, Xu JL, Li XZ, Zhang LH. AiiA, an enzyme that inactivates the acylhomoserine lactone quorum-sensing signal and attenuates the virulence of *Erwinia carotovora*. *Proc Natl Acad Sci USA*. 2000;97:3526–31.
- Carlier A, Uroz S, Smadja B, Fray R, Latour X, Dessaux Y, Faure D. The Ti plasmid of *Agrobacterium tumefaciens* harbors an *attM*-paralogous gene, *aiiB*, also encoding *N*-acyl homoserine lactonase activity. *Appl Environ Microbiol*. 2003;69:4989–93.
- Zhang HB, Wang LH, Zhang LH. Genetic control of quorum-sensing signal turnover in *Agrobacterium tumefaciens*. *Proc Natl Acad Sci USA*. 2002;99:4638–43.
- Uroz S, Oger PM, Chapel E, Adeline MT, Faure D, Dessaux Y. A *Rhodococcus* *qsdA*-encoded enzyme defines a novel class of large-spectrum quorum-quenching lactonases. *Appl Environ Microbiol*. 2008;74:1357–66.
- Lin YH, Xu JL, Hu J, Wang LH, Ong SL, Leadbetter JR, Zhang LH. Acyl-homoserine lactone acylase from *Ralstonia* strain XJ12B represents a novel and potent class of quorum-quenching enzymes. *Mol Microbiol*. 2003;47:849–60.
- Huang JJ, Han J, Zhang LH, Leadbetter JR. Utilization of acyl-homoserine lactone quorum signals for growth by a soil pseudomonad and *Pseudomonas aeruginosa* PAO1. *Appl Environ Microbiol*. 2003;69:5941–9.
- Park SY, Kang HO, Jang HS, Lee JK, Koo BT, Yum DY. Identification of extracellular *N*-acylhomoserine lactone acylase from a *Streptomyces* sp. and its application to quorum quenching. *Appl Environ Microbiol*. 2005;71:2632–41.
- Wong CS, Koh CL, Chan KG. Degradation of bacterial quorum sensing signaling molecules by the microscopical yeast *Trichosporon loubieri* isolated from tropical wetland waters. *Sensors*. 2013;13:12943–57.
- Yang F, Wang LH, Wang J, Dong YH, Hu JY, Zhang LH. Quorum quenching enzyme activity is widely conserved in the sera of mammalian species. *FEBS Lett*. 2005;579:3713–7.
- Schipper C, Hornung C, Bijtenhoorn P, Quitschau M, Grond S, Streit WR. Metagenome-derived clones encoding two novel lactonase family proteins involved in biofilm inhibition in *Pseudomonas aeruginosa*. *Appl Environ Microbiol*. 2009;75:224–33.
- Link AJ, Phillips D, Church GM. Methods for generating precise deletions and insertions in the genome of wild-type *Escherichia coli*: application to open reading frame characterization. *J Bacteriol*. 1997;179:6228–37.
- Manefield M, de Nys R, Naresh K, Roger R, Givskov M, Peter S, Kjelleberg S. Evidence that halogenated furanones from *Delisea pulchra* inhibit acylated homoserine lactone (AHL)-mediated gene expression by displacing the AHL signal from its receptor protein. *Microbiology*. 1999;145:283–91.
- Karam G, Chastre J, Wilcox MH, Vincent JL. Antibiotic strategies in the era of multidrug resistance. *Crit Care*. 2016;20:136.
- Tang KH, Zhang XH. Quorum quenching agents: resources for antiviral therapy. *Mar Drugs*. 2014;12:3245–82.
- Lee S, Park SK, Kwon H, Lee SH, Lee K, Nahm CH, Jo SJ, Oh HS, Park PK, Choo KH, Lee CH, Yi T. Crossing the border between laboratory and field: bacterial quorum quenching for anti-biofouling strategy in an MBR. *Environ Sci Technol*. 2016;50:1788–95.
- Oh HS, Tan CH, Low JH, Rzechowicz M, Siddiqui MF, Winters H, Kjelleberg S, Fane AG, Rice SA. Quorum quenching bacteria can be used to inhibit the biofouling of reverse osmosis membranes. *Water Res*. 2017;112:29–37.
- Díaz E, Jiménez JJ, Nogales J. Aerobic degradation of aromatic compounds. *Curr Opin Biotechnol*. 2013;24:431–42.
- Yong YC, Zhong JJ. *N*-acylated homoserine lactone production and involvement in the biodegradation of aromatics by an environmental isolate of *Pseudomonas aeruginosa*. *Process Biochem*. 2010;45:1944–8.
- See-Too WS, Ee R, Lim YL, Convey P, Pearce DA, Yin WF, Chan KG. *AidP*, a novel *N*-acyl homoserine lactonase gene from Antarctic *Planococcus* sp. *Sci Rep*. 2017;7:42968.
- See-Too WS, Ee R, Madhaiyan M, Kwon SW, Tan JY, Lim YL, Convey P, Pearce D, Yin WF, Chan KG. *Planococcus versusus* sp. nov., isolated from soil. *Int J Syst Evol Microbiol*. 2016;67:944–50.
- McClellan K, Winson M, Fish L, Taylor A, Chhabra S, Camara M, Daykin M, Lamb J, Swift S, Bycroft B, Stewart G, Williams P. Quorum sensing and *Chromobacterium violaceum*: exploitation of violacein production and inhibition for the detection of *N*-acylhomoserine lactones. *Microbiology*. 1997;143:3703–11.
- Kumar S, Stecher G, Tamura KMEGA. 7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Mol Biol Evol*. 2016;33:1870–4.
- Edgar RC. Muscle: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res*. 2004;32:1792–7.
- Waterhouse AM, Procter JB, Martin DM, Clamp M, Barton GJ. Jalview version 2-A multiple sequence alignment editor and analysis workbench. *Bioinformatics*. 2009;25:1189–91.
- Yang Z. PAML 4: phylogenetic analysis by maximum likelihood. *Mol Biol Evol*. 2007;24:1586–91.
- Yang Z, Wong WS, Nielsen R. Bayes empirical Bayes inference of amino acid sites under positive selection. *Mol Biol Evol*. 2005;22:1107–18.
- Biasini M, Bienert S, Waterhouse A, Arnold K, Studer G, Schmidt T, Kiefer F, Gallo Cassarino T, Bertoni M, Bordoli L, Schwede T. SWISS-MODEL: modelling protein tertiary and quaternary structure using evolutionary information. *Nucleic Acids Res*. 2014;42:252–8.
- Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem*. 1976;72:248–54.
- Todd MJ, Gomez Z. Enzyme kinetics determined using calorimetry: a general assay for enzyme activity? *Anal Biochem*. 2001;296:179–87.
- Lojkowska E, Masclaux C, Boccarda M, Robert-Baudouy J, Hugouvieux-Cotte-Pattat N. Characterization of the *pelL* gene encoding a novel peptidate lyase of *Erwinia chrysanthemi* 3937. *Mol Microbiol*. 1995;16:1183–95.
- Uroz S, D'Angelo-Picard C, Carlier A, Elasmri M, Sicot C, Petit A, Oger P, Faure D, Dessaux Y. Novel bacteria degrading *N*-acylhomoserine lactones and their use as quenchers of quorum-sensing-regulated functions of plant-pathogenic bacteria. *Microbiology*. 2003;149:1981–9.
- Liu D, Momb J, Thomas PW, Moulin A, Petsko GA, Fast W, Ringe D. Mechanism of the quorum-quenching lactonase (AiiA) from *Bacillus thuringiensis*. 1. Product-bound structures. *Biochemistry*. 2008;47:7706–14.
- Momb J, Wang C, Liu D, Thomas PW, Petsko GA, Guo H, Ringe D, Fast W. Mechanism of the quorum-quenching lactonase (AiiA) from *Bacillus*

- thuringiensis*. 2. Substrate modeling and active site mutations. *Biochemistry*. 2008;47:7715–25.
47. Aghajari N, Feller G, Gerday C, Haser R. Structures of the psychrophilic *Alteromonas haloplanctis* α -amylase give insights into cold adaptation at a molecular level. *Structure*. 1998;6:1503–16.
 48. DasSarma S, Capes MD, Karan R, DasSarma P. Amino acid substitutions in cold-adapted proteins from *Halorubrum lacusprofundi*, an extremely halophilic microbe from Antarctica. *PLoS ONE*. 2013;8:e58587.
 49. Feller G. Protein stability and enzyme activity at extreme biological temperatures. *J of Phys*. 2010;22:32.
 50. Tattersall GJ, Sinclair BJ, Withers PC, Fields PA, Seebacher F, Cooper CE, et al. Coping with thermal challenges: physiological adaptations to environmental temperatures. *Compr Physiol*. 2012;2:2151–202.
 51. Mavromatis K, Tsigos I, Tzanodaskalaki M, Kokkinidis M, Bouriotis V. Exploring the role of a glycine cluster in cold adaptation of an alkaline phosphatase. *Eur J Biochem*. 2002;269:2330–5.
 52. Feller G, Payan F, Theys F, Qian M, Haser R, Gerday C. Stability and structural analysis of α -amylase from the antarctic psychrophile *Alteromonas haloplanctis* A23. *Eur J Biochem*. 1994;222:441–7.
 53. Davail S, Feller G, Narinx E, Gerday C. Cold adaptation of proteins. Purification, characterization, and sequence of the heat-labile subtilin from the Antarctic psychrophile *Bacillus* TA41. *J Biol Chem*. 1994;269:17448–53.
 54. Hawwa R, Aikens J, Turner R, Santarsiero BD, Mesecar AD. Structural basis for thermostability revealed through the identification and characterization of a highly thermostable phosphotriesterase-like lactonase from *Geobacillus stearothermophilus*. *Arch Biochem Biophys*. 2009;488:109–20.
 55. Yancey PH, Somero GN. Temperature-dependence of intracellular pH—its role in conservation of pyruvate apparent K_m values of vertebrate lactate-dehydrogenases. *J Comp Physiol Psychol*. 1978;125:129–34.
 56. Momb J, Thomas PW, Breece RM, Tierney DL, Fast W. The quorum-quenching metallo- γ -lactonase from *Bacillus thuringiensis* exhibits a leaving group trio effect. *Biochemistry*. 2006;45:13385–93.
 57. Saavedra HG, Wrabl JO, Anderson JA, Li J, Hilsner VJ. Dynamic allostery can drive cold adaptation in enzymes. *Nature*. 2018;558:324–8.
 58. Tang K, Su Y, Brackman G, Cui F, Zhang Y, Shi X, Coenye T, Zhang XH. MomL, a novel marine-derived *N*-acyl homoserine lactonase from *Muricauda olearia*. *Appl Environ Microbiol*. 2015;81:774–82.
 59. Wang WZ, Morohoshi T, Someya N, Ikeda T. AidC, a novel *N*-acylhomoserine lactonase from the potato root-associated cytophaga-flavobacteriobacteroides (CFB) group bacterium *Chryseobacterium* sp. strain StRB126. *Appl Environ Microbiol*. 2012;78(22):7985–92.
 60. Dong W, Zhu J, Guo X, Kong D, Zhang Q, Zhou Y, Liu X, Zhao S, Ruan Z. Characterization of AiiK, an AHL lactonase, from *Kurthia huakui* LAM0618^T and its application in quorum quenching on *Pseudomonas aeruginosa* PAO1. *Sci Rep*. 8. Article number: 6013.
 61. Bergonzi C, Schwab M, Elias M. The quorum-quenching lactonase from *Geobacillus caldoolysilyticus*: purification, characterization, crystallization and crystallographic analysis. *Acta Crystallogr F Struct Biol Commun*. 2016;72:681–6.
 62. Morohoshi T, Tominaga Y, Someya N, Ikeda T. Characterization of a novel thermostable *N*-acylhomoserine lactonase from the thermophilic bacterium *Thermaerobacter marianensis*. *J Biosci Bioeng*. 2015;120:1–5.
 63. Wang LH, Weng LX, Dong YH, Zhang LH. Specificity and enzyme kinetics of the quorum-quenching *N*-acyl homoserine lactone lactonase (AHL-lactonase). *J Biol Chem*. 2004;279:13645–51.
 64. Liu D, Thomas PW, Momb J, Hoang QQ, Petsko GA, Ringe D, Fast W. Structure and specificity of a quorum-quenching lactonase (AiiB) from *Agrobacterium tumefaciens*. *Biochemistry*. 2007;46:11789–99.
 65. Garge SS, Nerurkar AS. Attenuation of quorum sensing regulated virulence of *Pectobacterium carotovorum* subsp. *carotovorum* through an AHL lactonase produced by *Lysinibacillus* sp. G550. *PLoS ONE*. 2016;11:e0167344.
 66. Migiyama Y, Kaneko Y, Yanagihara K, Morohoshi T, Morinaga Y, Nakamura S, et al. Efficacy of AiiM, an *N*-acylhomoserine lactonase, against *Pseudomonas aeruginosa* in a mouse model of acute pneumonia. *Antimicrob Agents Chemother*. 2013;57:3653–8.
 67. Morohoshi T, Tominaga Y, Someya N, Ikeda T. Complete genome sequence and characterization of the *N*-acylhomoserine lactone-degrading gene of the potato leaf-associated *Solibacillus silvestris*. *J Biosci Bioeng*. 2012;113:20–5.
 68. Mei GY, Yan XX, Turak A, Luo ZQ, Zhang LQ. AidH, an α / β -hydrolase fold family member from an *Ochrobactrum* sp. strain, is a novel *N*-acylhomoserine lactonase. *Appl Environ Microbiol*. 2010;76:4933–4.
 69. Torres M, Uroz S, Salto R, Fauchery L, Quesada E, Llamas I, HqiA, a novel quorum-quenching enzyme which expands the AHL lactonase family. *Sci Rep*. 2017;7:943.
 70. Dong YH, Wang L, Xu JL, Zhang HB, Zhang XF, Zhang LH. Quenching quorum-sensing-dependent bacterial infection by an *N*-acyl homoserine lactonase. *Nature*. 2001;411:813–7.
 71. García-Contreras R, Maeda T, Wood TK. Can resistance against quorum-sensing interference be selected? *ISME J*. 2016;10:4–10.
 72. Chen F, Gao Y, Chen X, Yu Z, Li X. Quorum quenching enzymes and their application in degrading signal molecules to block quorum sensing-dependent infection. *Int J Mol Sci*. 2013;14:17477–500.
 73. Schikora A, Schenk ST, Stein E, Molitor A, Zuccaro A, Kogel KH. *N*-acylhomoserine lactone confers resistance toward biotrophic and hemibiotrophic pathogens via altered activation of atmpk6. *Plant Physiol*. 2011;157:1407–18.
 74. Bai X, Todd CD, Desikan R, Yang Y, Hu X. *N*-3-oxo-decanoyl-H-homoserine-lactone activates auxin-induced adventitious root formation via hydrogen peroxide- and nitric oxide-dependent cyclic gmp signaling in mung bean. *Plant Physiol*. 2012;158:725–36.
 75. Veliz-Vallejos DF, van Noorden GE, Yuan M, Mathesius U. A *Sinorhizobium meliloti*-specific *N*-acyl homoserine lactone quorum sensing signal increases nodule numbers in *Medicago truncatula* independent of autoregulation. *Front Plant Sci*. 2014;5:551.
 76. Ali S. Plant growth-promoting bacterial endophytes that contain acc deaminase: isolation, characterization, and use; 2013. Thesis, University of Waterloo, Ontario, Canada.
 77. Sessitsch A, Coenye T, Sturz AV, Vandamme P, Barka EA, Salles JF, Van Elsas JD, Faure D, Reiter B, Glick BR, Wang-Pruski G, Nowak J. *Burkholderia phytofirmans* sp. nov., a novel plant-associated bacterium with plant-beneficial properties. *Int J Syst Evol Microbiol*. 2005;55:1187–92.
 78. Liu X, Jia J, Popat R, Ortori CA, Li J, Diggle SP, Gao K. Characterisation of two quorum sensing systems in the endophytic *Serratia plymuthica* strain G3: differential control of motility and biofilm formation according to life-style. *BMC Microbiol*. 2011;11:26.